

# Outils bio-informatiques pour la protéomique- Comparaison TheGPM/Xcalibur Sequest- Gelprint: génération de gels 2D in-silico

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# De quoi parlerons nous?

- Présentation de TheGPM
- Mimivirus: un virus géant vivant dans les amibes
- Comparaison TheGPM/Sequest pour l'identification des protéines par spectrométrie de masse MS/MS

The GPM - Mozilla

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 The Global Proteome Machine Organization  
www.thegpm.org

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**Single proteomes**

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- [T. brucei](#)

**Projects**

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- [LiveCD](#)
- [Quartz](#)
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**GPM News**

**Two new Projects available: LiveCD and Quartz (2004/4/15)**

The GPMO has added two new projects, LiveCD and Quartz to the site. LiveCD, a project from the University of Michigan NCRR Center for Proteomics, provides a simple method to install a Linux-based version of X! TANDEM and the GPM on a large number of computers for instructional and demonstration purposes. It also includes some software allowing the use of X! TANDEM on clusters of computers running LiveCD.

Quartz is a GPMO staff project. It is a set of annotated spectrum collections, meant to be used for bioinformatics research. The current collections contain > 2000 MS/MS spectra, along with XML-formatted annotation files.

**X! TANDEM and the GPM release updates (2004/4/10)**

New releases of both X! TANDEM and the GPM were released today. This is a maintenance release, including fixes for small problems observed with previous versions. The collections of sequences for the GPM have been updated to include the latest sequence releases from ENSEMBL (1/4/2004).

**Probit model published (2004/3/1)**

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### The Global Proteome Machine



**spectra:** DTA, PKL or Matrix Science format only

**taxon:** Select the appropriate species.

Homo sapiens  
Mus musculus  
Rattus norvegicus  
D. melanogaster  
D. rerio  
C. elegans  
A. thaliana  
S. cerevisiae

**fragment  $\delta m$ :**  Da  ppm

**output:** log(e) <

**modifications:** format = "m<sub>1</sub>@X<sub>1</sub>,m<sub>2</sub>@X<sub>2</sub>..." where:  
m<sub>i</sub> = mod. mass (Da) & X<sub>i</sub> = residue.

complete:   
potential:   
potential:   
(refine)  
motif:   
(refine)

**mutations:**  yes  no

**method:** Select device & parent  $\delta m$ .

FTICR (10 ppm)  
Quad-TOF (100 ppm)  
Quad-TOF (0.5 Da)  
Ion Trap (4 Da)

Par défaut GPM est configuré pour utiliser les protéomes suivants:

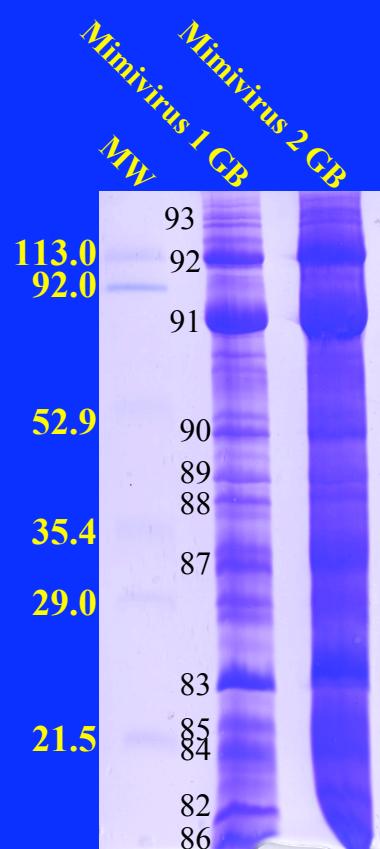
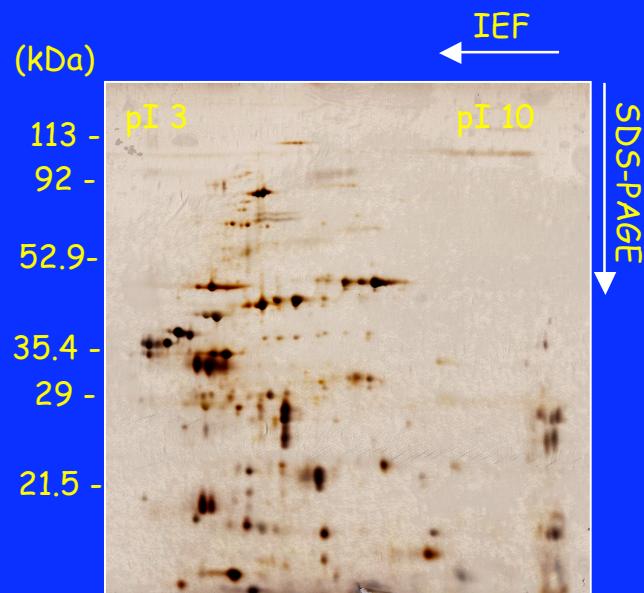
1. Human (34d NCBI 34, 10 May 2004)
2. Mouse (32b NCBI m32, 1 Apr. 2004)
3. Rat (RGSC 3.1b, 9 Feb. 2004)
4. Zebra fish (3b WTSI Zv3, 1 Apr. 2004)
5. Fruit fly (BGDP 3.1, 2 Jul. 2003)
6. C. elegans (116a WS 116, 1 Apr. 2004)
7. Yeast (SCD, 15 Nov 2003)
8. A. thaliana (ATH1, v. 5.0, Jan 29, 2004)

Et fonctionne sur une machine standard, sous Windows ou Linux:  
(nous avons teste linux)

1. 2.4 GHz Pentium IV processors;
2. 256 MB memory
3. 40 GB, 7200 rpm EIDE hard drive
4. Apache HTTP server, v. 2
5. ActiveState Perl

Notre expérience est basée sur des spots issu de gels:

- 2D coloré au nitrate d'argent
- 1 D coloré au bleu de Coomassie



Spots analysés sur une trappe à ions LCQ Deca XP plus

the GPM site - Mozilla

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### The Global Proteome Machine



**spectra:** DTA, PKL or Matrix Science format only  
 [Browse...](#)

**taxon:** Select the appropriate species.

D. melanogaster  
D. rerio  
C. elegans  
A. thaliana  
S. cerevisiae  
Mimivirus prots  
Swissprot May 2004  
trEMBL May 2004

**Find models**

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[north america](#)

| [h003](#) | [h066](#) | [h112](#) |  
| [h319](#) | [h451](#) | [h777](#) |  
| [h874](#) |

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**fragment  $\delta m$ :**   Da  ppm

**output:**  $\log(e) <$

**modifications:** format = "m<sub>1</sub>@X<sub>1</sub>,m<sub>2</sub>@X<sub>2</sub>..." where:  
m<sub>i</sub> = mod. mass (Da) & X<sub>i</sub> = residue.

complete:   
potential:   
refinement:

---

**mutations:**  yes  no

**method:** Select device & parent  $\delta m$ .

FTICR (10 ppm)  
Quad-TOF (100 ppm)  
Quad-TOF (0.5 Da)  
Ion Trap (4 Da)

[... view method](#)

the GPM site - Mozilla

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**The Global Proteome Machine**  
advanced search page



**spectra:** DTA, PKL or Matrix Science format only

**taxon:** Mimivirus prots

**measurement errors**

Fragment mass error: 0.4 Da

Parent mass error: +100 100 ppm

Isotope error:  yes  no

**output**

Max expect <: 0.1

**residue modifications**

modifications: 57.01@C  
m1@X,m2@Y, etc.

**potential modifications**

16@M  
m1@X,m2@Y, etc.

protein N-terminus: 0.0 Da

protein C-terminus: 0.0 Da

**cleavage specification**

cleavage C-terminal change: +17.00305 Da

the GPM site - Mozilla

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cleavage site:  trypsin = [KR]|{P}

missed sites:

**model refinement**

refine model:  yes  no

point mutations:  yes  no

potential modifications:   
m1@X,m2@Y, etc.

use these modifications throughout:  yes  no

unanticipated cleaves ([X][X]):  yes  no

potential N-terminus modifications:

potential C-terminus modifications:

valid expectation: <

**spectrum synthesis**

spectrum synthesis:  yes  no

**spectrum conditioning**

Noise suppression:  yes  no

Minimum parent M+H:  Da

Minimum fragment m/z:

Total peaks:

Minimum peaks:

GPM - Models from 'bc91.mgf' - Mozilla

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Models from 'bc91.mgf'

[PERFORMANCE](#) | [parameters](#) | [details](#) | [XML](#) |

[get annotation](#)

rank	log(e)	accession	
#1.	-125.2	Contig31_171[63834-62038](REVERSESENSE)	<a href="#">homologues protein</a>
#2.	-73.8	Contig31_180[46236-44239](REVERSESENSE)	<a href="#">homologues protein</a>
#3.	-7.0	Contig31_115[196187-194196](REVERSESENSE)	<a href="#">homologues protein</a>
#4.	-3.9	Contig33_76[165386-167494]	<a href="#">homologues protein</a>
#5.	-2.7	gi 999627	<a href="#">homologues protein</a>
		Chain B, Porcine E-Trypsin (E.C.3.4.21.4).	
#6.	1.7	Contig32_164[18449-13218](REVERSESENSE)	<a href="#">homologues protein</a>

plist.pl, v. 2004.03.01

Major capsid protein



GPM - protein model: Contig31\_171[63834-62038](REVERSESENSE) - Mozilla

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protein model: Contig31\_171[63834-62038](REVERSESENSE)

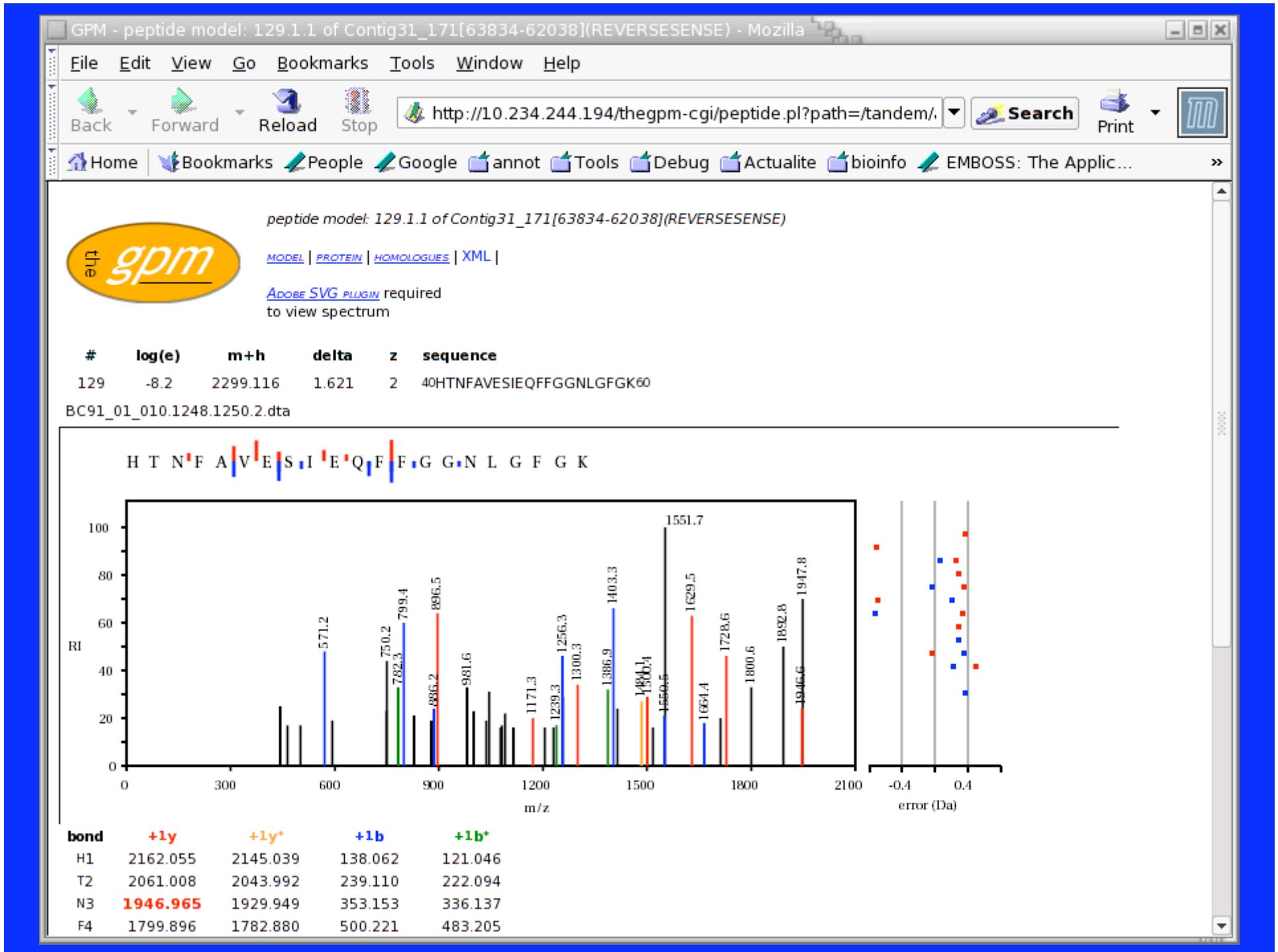
[MODEL](#) | [homologues](#) | [details](#) | [XML](#) |

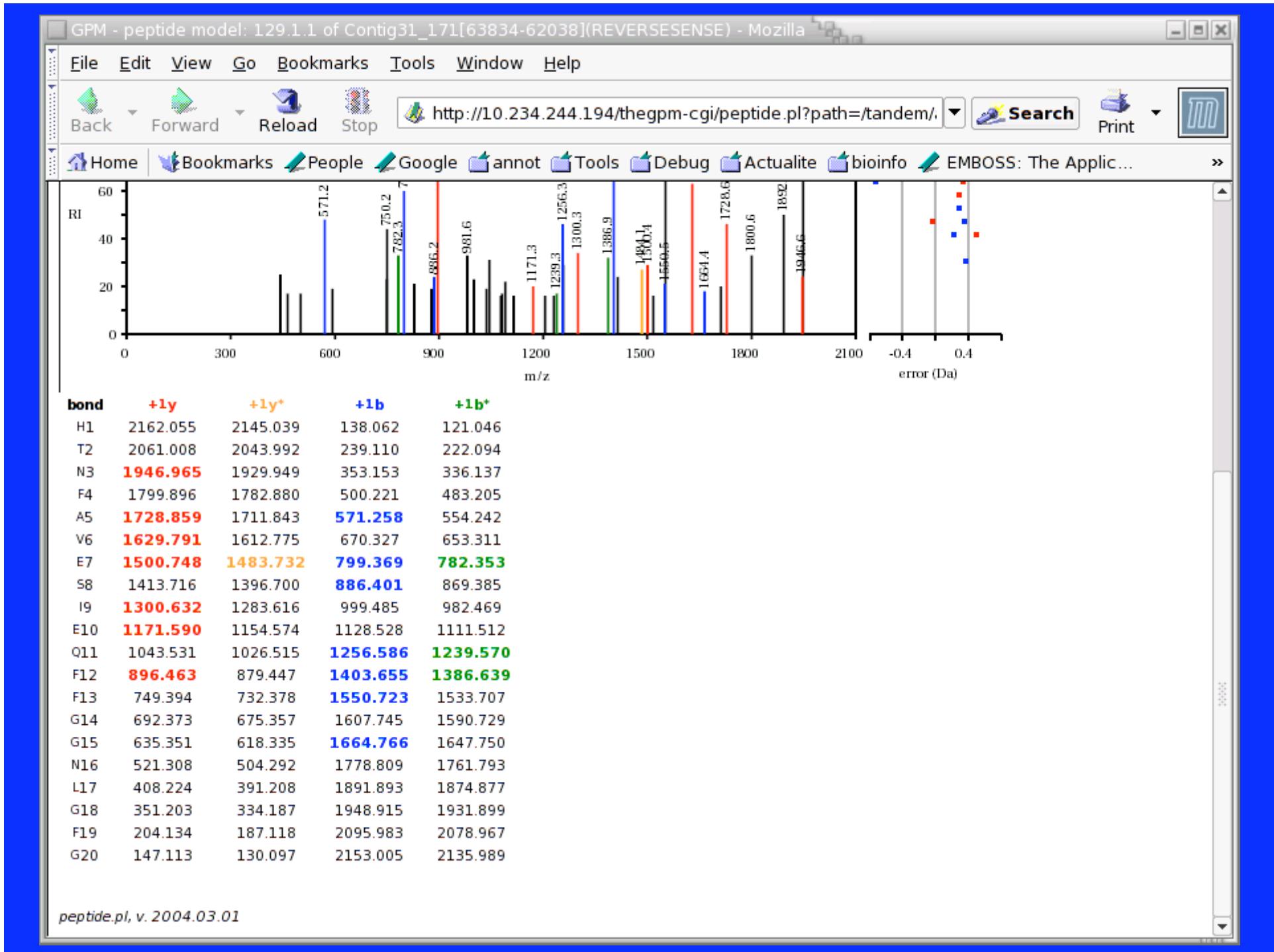
ensembl

**log(e) = -125.2** Contig31\_171[63834-62038](REVERSESENSE)

1 VTSSTTGRLWETKFLKVQSSPLGKLRGNRVFKVVYRR**HTNFAVESIEQFFGGNLGFGK** 60  
61 KSSAEINR**SGDLITQVFLKVTLP**EVRYCGDFTNFGHVEFAWVR**NIGHAI**VEETELEIGGS 120  
121 **PIDKH**YGDWLQIWQDVSSSKDHEKGLAKMLGDVPELTSISTLSWDVPDNTVLKPSYTLYV 180  
181 PLQFYFNR**NNGLALPLIALQYHQVR**IYVKFR**QADQCYIASDAFK**SGCGNLQLDDVSLVN 240  
241 YVFLDTEERRRFAQVSHEYLIEQLQFTGEESAGSSNSAKYKLNFNHPVKAIYWVTK**LGNY** 300  
301 **QGGKFMTYDPVCWENAR**ENAAKLLLLAQYDLDWDGYFQEPEGGYECEGNDGRSYVGDCGVQ 360  
361 YTAVDPSNPSEEPSYIFNDTTAEEFDGSLLIGK**LAPCVPLLKR**RNKDVDLKD**VEGIIRI** 420  
421 **HTDFENDRMKYPEVEK**ITR**NDLTLHDLSVPISKY**DVDNRVDYIKKFDTVWQHNNFGLLI 480  
481 DGSGNPTHEAELQLNGQPRQSKR**GGI**WYDTVNPTVHHTKSPRDGVNVFSFALNPEEHQPS 540  
541 **CTCNFSRIDTAQLNLWFQHFTNHKFADVFADNDNKVLIFAV**NYNVLRMLSGMAGLAYSN 599

spectrum	log(e)	m+h	delta	z	sequence
129.1	-8.2	2299.116	1.621	2	vyrr40 <b>HTNFAVESIEQFFGGNLGFGK</b> 60kssa
220.1	-6.1	2299.116	0.696	2	vyrr40 <b>HTNFAVESIEQFFGGNLGFGK</b> 60kssa
128.1	-3.7	2300.116	1.681	3	vyrr40 <b>HTNFAVESIEQFFGGNLGFGK</b> 60kssa
210.1	-2.9	2300.116	0.677	3	vyrr40 <b>HTNFAVESIEQFFGGNLGFGK</b> 60kssa
57.1	-5.6	1220.690	0.467	2	einr69 <b>SGDLITQVFLK</b> 79vtlp
1087.1	-4.7	1221.690	0.902	2	einr69 <b>SGDLITQVFLK</b> 79vtlp





GPM - spectra view - Mozilla

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main spectra listing

 MODEL

#129, e = 5.8e-09, M+H = 22300.741.621, VYRR<sub>40</sub>HTNFAVESIEQFFGGNLGFGK<sub>60</sub>KSSA,  
**log(E) = -125.2, Contig31\_171**  
Model protein sequences  
Supporting evidence

#128, e = 1.9e-04, M+H = 32301.81.681, VYRR<sub>40</sub>HTNFAVESIEQFFGGNLGFGK<sub>60</sub>KSSA,  
**log(E) = -125.2, Contig31\_171**  
Model protein sequences  
Supporting evidence

#210, e = 1.4e-03, M+H = 32300.790.677, VYRR<sub>40</sub>HTNFAVESIEQFFGGNLGFGK<sub>60</sub>KSSA,  
**log(E) = -125.2, Contig31\_171**  
Model protein sequences  
Supporting evidence

#220, e = 7.6e-07, M+H = 22299.810.696, VYRR<sub>40</sub>HTNFAVESIEQFFGGNLGFGK<sub>60</sub>KSSA,  
**log(E) = -125.2, Contig31\_171**  
Model protein sequences  
Supporting evidence

#1081, e = 4.5e-02, M+H = 1979.6790.168, EINR<sub>69</sub>SGDLITQVF<sub>77</sub>LKVT,  
**log(E) = -125.2, Contig31\_171**  
Model protein sequences  
Supporting evidence

#135, e = 9.1e-05, M+H = 21222.560.875, EINR<sub>69</sub>SGDLITQVFLK<sub>79</sub>VTL<sub>P</sub>,  
**log(E) = -125.2, Contig31\_171**  
Model protein sequences  
Supporting evidence

#57, e = 2.7e-06, M+H = 21221.160.467, EINR<sub>69</sub>SGDLITQVFLK<sub>79</sub>VTL<sub>P</sub>,  
**log(E) = -125.2, Contig31\_171**

GPM - performance - Mozilla

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 modelling performance statistics

[MODEL](#) | [parameters](#) | [XML](#) | [spectra view](#)

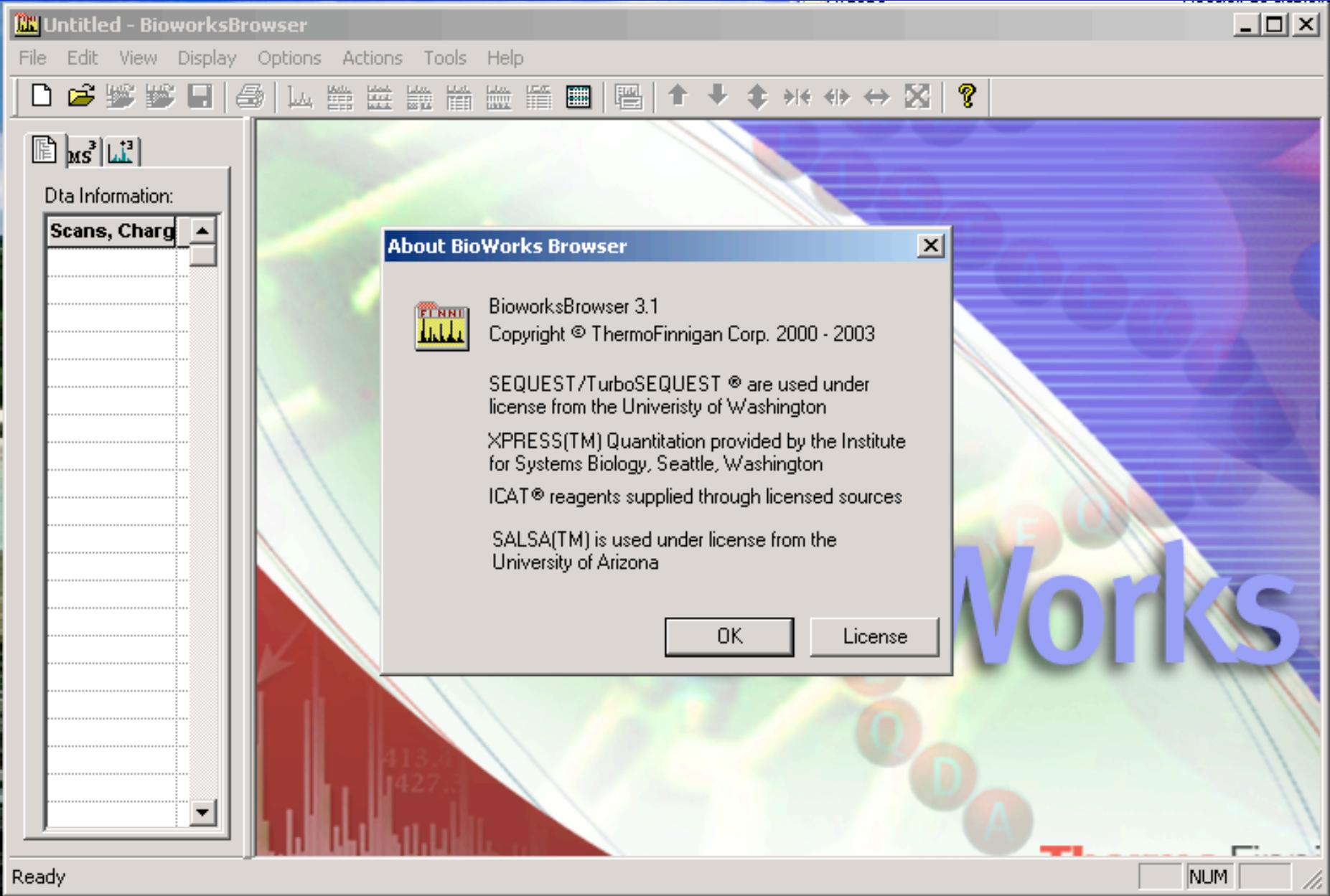
**Performance statistics:**

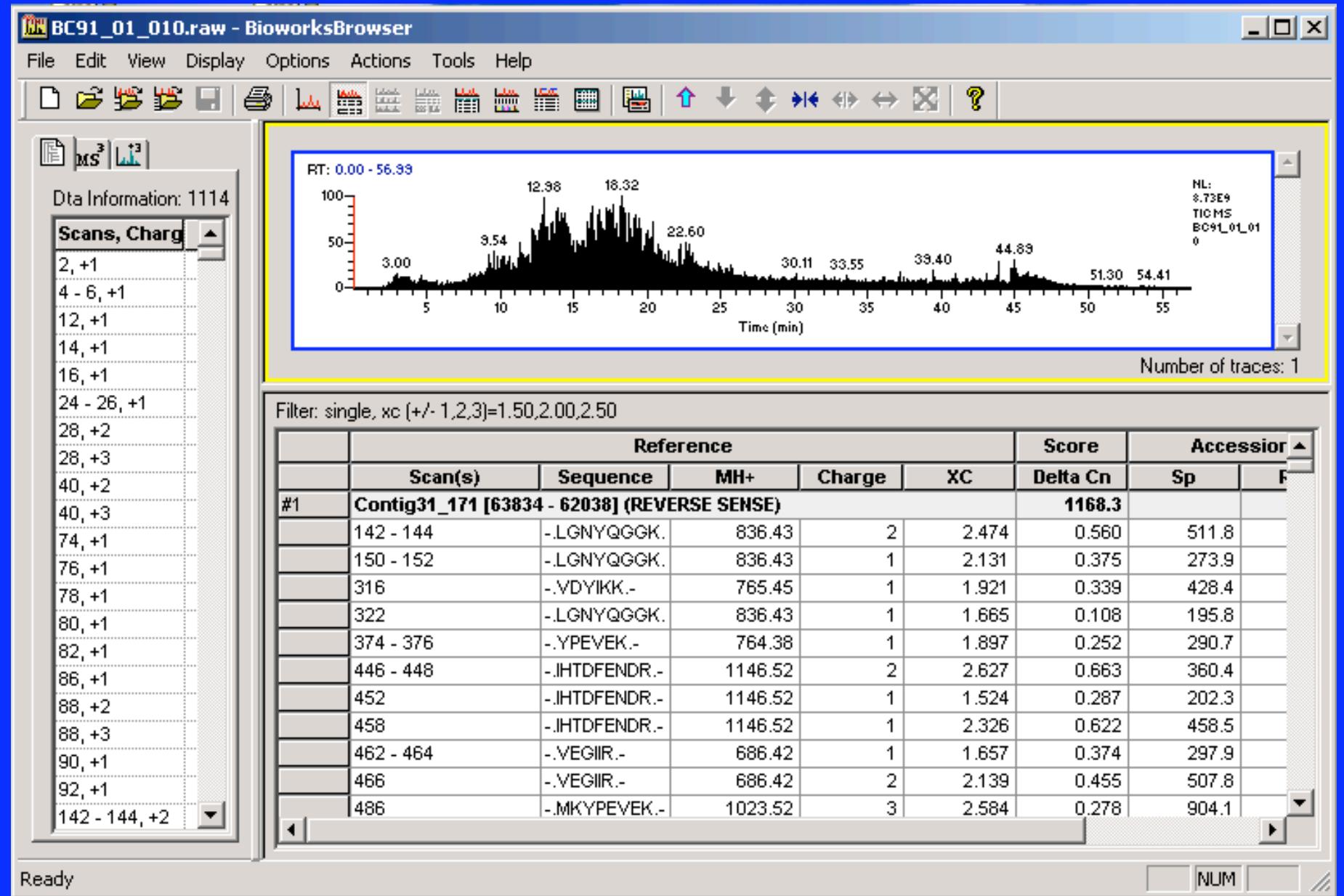
Parameter	Value
list path, sequence source #1	..fasta/mimi.fasta
list path, sequence source #2	..fasta/crap.fasta.pro
modelling, spectrum noise suppression ratio	0.09
modelling, total proteins used	1259
modelling, total spectra assigned	139
modelling, total spectra used	1114
process, start time	2004:05:26:16:51:12
process, version	x! tandem 2004.04.01
refining, # input models	12
refining, # input spectra	899
refining, # partial cleavage	11
refining, # potential N-terminii	2
refining, # unanticipated cleavage	26
timing, initial modelling total (sec)	1.10
timing, initial modelling/spectrum (sec)	0.001
timing, load sequence models (sec)	0.02
timing, refinement/spectrum (sec)	0.003

perform.pl, v. 2004.03.01

1200 séquences protéiques dans la base de données  
1114 spectres au départ

Temps: 1.10s





**Coverage - Contig31\_171 [63834 - 62038] (REVERSE SENSE)**

Reference: Contig31\_171 [63834 - 62038] (REVERSE SENSE)

Database: mimi.fasta      Monoisotopic Mass: 68033.9      Number of Amino Acids: 599      pI: 5.62

	1-10	11-20	21-30	31-40	41-50	51-60	61-70	71-80	81-90	91-100
1	VTSSTTGRLW	VETKFLLKVQ	SSPLGKLRGN	RVFKVVYRRH	TNFIAVE SIEQ	FFGGNLGFGK	KSSAEINRSG	DLITQVFLKV	TLPEVRYCGD	FTNFGHVFA
101	WVRNIGHAIW	EETELEIGGS	PIDKHYGDWL	QIWQDVSSSK	DHEKGLAKML	GDVPELTSIS	TLSWDVPDNT	VLKPSYTLYV	PLQFYFMRNN	GLALPLIALQ
201	YHQVRIYVKF	RQADQCYIAS	DAFKSGCGNL	Q1DDWSLYVN	YVFLDTERR	RFAQVSHEYL	IEQLQFTGEE	SAGSSNSAKY	KLNFNHPVKA	IYWVTKLGNY
301	QGGKFMITYDP	VCWENARENA	AKLLLALQYD	LDDWGYFQEP	GGYECEGNDG	RSYVGDCGVQ	YTAVDPSNPS	EEPSYIFNDT	TTAAEAFD GSL	LIGKLAPCVP
401	LLKRNKDVLDL	KDKVEGIIRI	HTDFENDRMK	YPEVEK ITRN	DLTLHDLSVP	ISKYDVDNRV	DYIKKFDTVTV	WQHNNFGLLI	DGSGNPTHEA	ELQLNGQPRQ
501	SKRGGIWYDT	VMPTVHHTKS	PRDGVNVFSF	ALNPEEHQPS	CTCNFSRIDT	AQLNLWFQHF	TNHKFADVFA	DNDNKVLIFA	VNYNVLRMLS	GMAGLAYSN

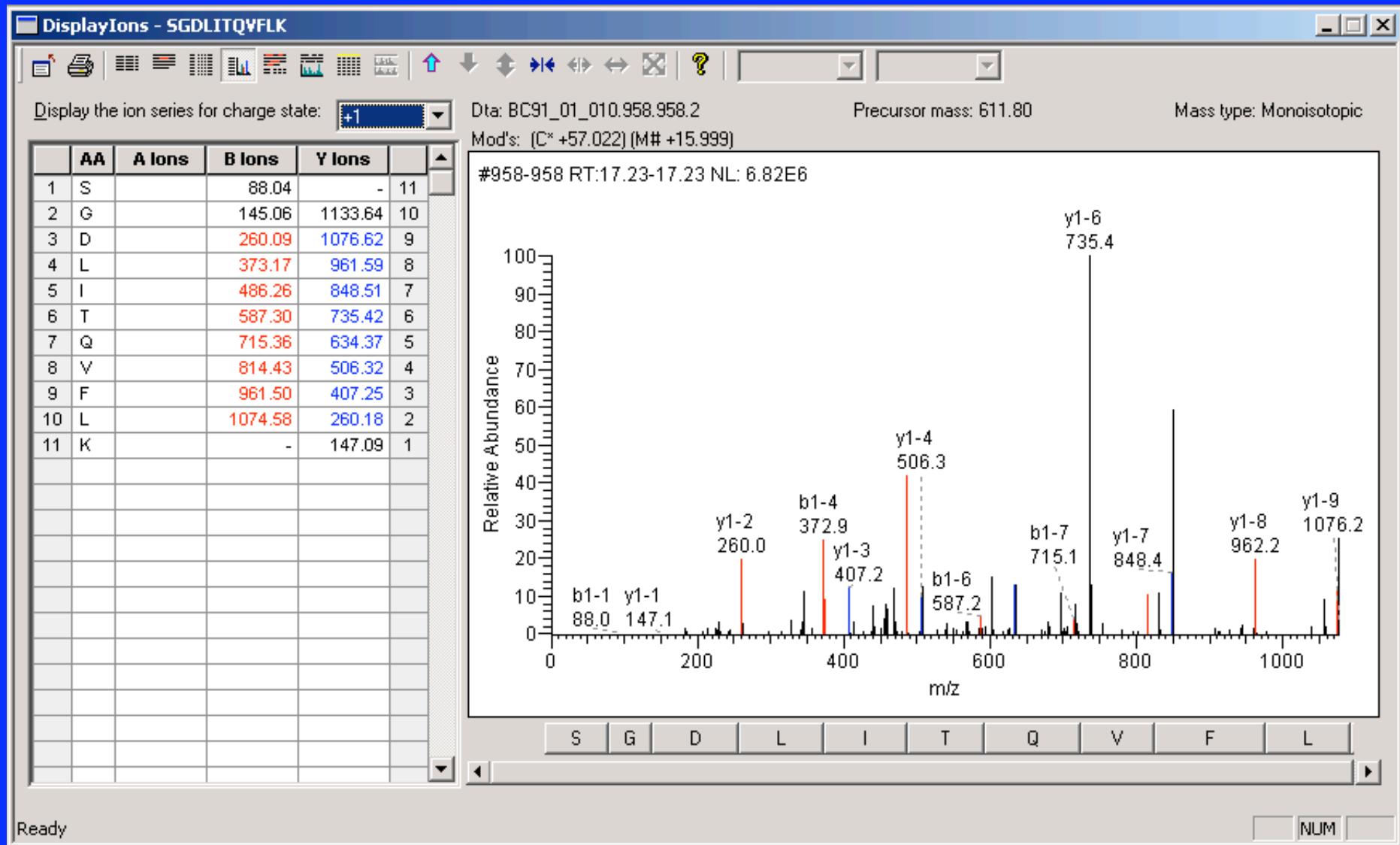
Protein Coverage:

	Sequence	MH+	% by Mass	Position	% by AA's
<input checked="" type="checkbox"/>	HTNFAVESIEQFFGGNLGFGK	2299.11	3.38	40 - 60	3.51
<input checked="" type="checkbox"/>	SGDLITQVFLK	1220.69	1.79	69 - 79	1.84
<input type="checkbox"/>	VTLP EVR	813.48	1.20	80 - 86	1.17
<input type="checkbox"/>	YCGDFTNFGHVFAWVR	2047.91	3.01	87 - 103	2.84
<input checked="" type="checkbox"/>	NIGHAIVEETELEIGGSPIDK	2221.14	3.26	104 - 124	3.51
<input checked="" type="checkbox"/>	NIGHAIVEETELEIGGSPIDK	2221.14	3.26	104 - 124	3.51
<input checked="" type="checkbox"/>	NIGHAIVEETELEIGGSPIDK	2221.14	3.26	104 - 124	3.51
<input type="checkbox"/>	NIGHAIVEETELEIGGSPIDK	2221.14	3.26	104 - 124	3.51
<input checked="" type="checkbox"/>	HYGDWLQM/QDVSSSK	1948.92	2.86	125 - 140	2.67

Protein Coverage Totals

by Mass:	27075.6
% by Mass:	39.80
by Position:	236
% by AA's:	39.40

Ready



Sur 10 échantillons,  
8 identifications  
cohérentes

Echantillon	Id match	Score gpm	Score sequest
82	-	-	-
83	29_13 34_197 34_273	- - -22.9	30 20 -
84	34_129 34_156	-19.9 -16.1	190 40
85	34_69 34_129	-14.7 -3.5	100 20
86	29_35	-13.9	30.2
87	31_171 31_180 31_7 31_161	-56.1 -26.6 -16.1 -13.1	300 110 70 20
88	31_161 34_103 34_288	-49 -23.7 -13.8	348 72 30
89	31_161 34_152	-50 -5.7	210 30
90	31_161	-111	1140
91	31_171 31_180 31_115	-125.2 -73.8 -7	1168 290 30

Conclusion:

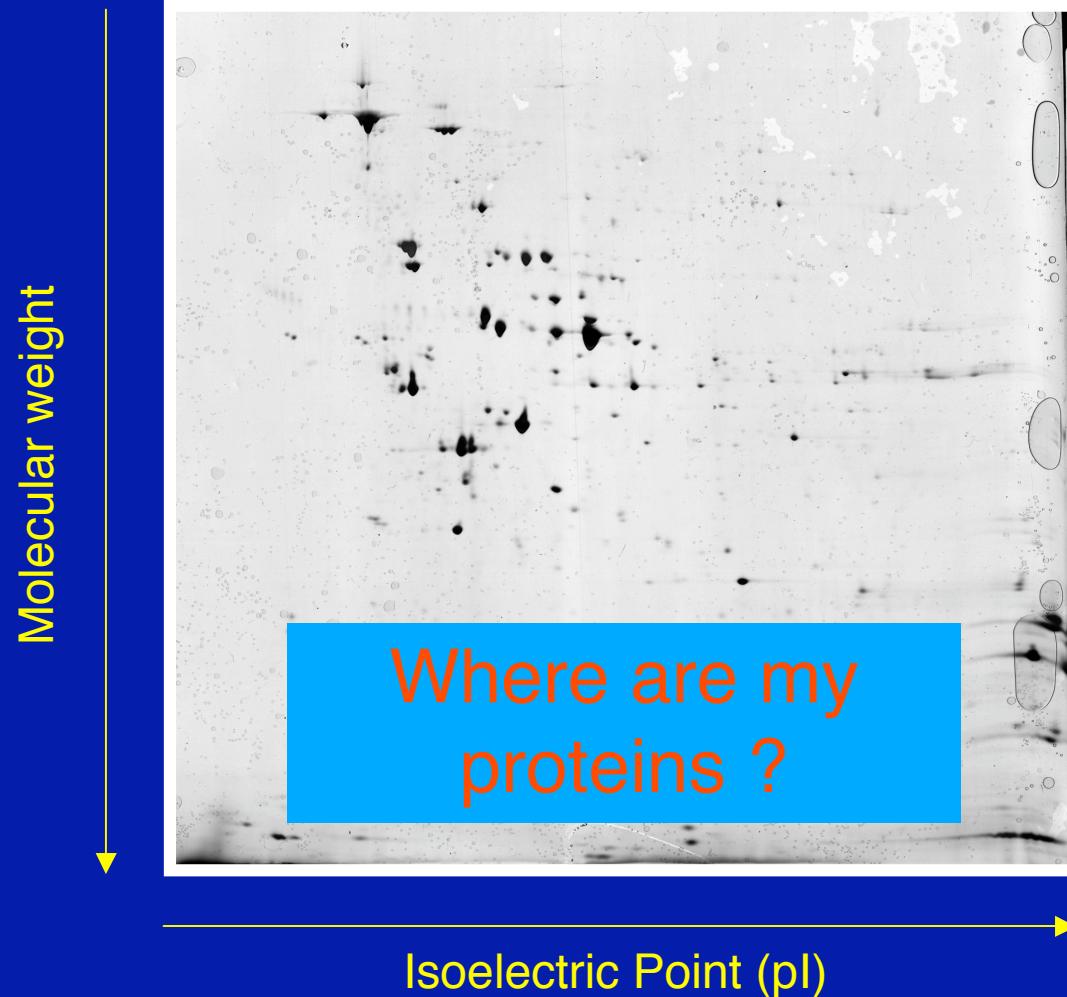
Une différence considérable en temps de calcul:

theGPM: 1 s

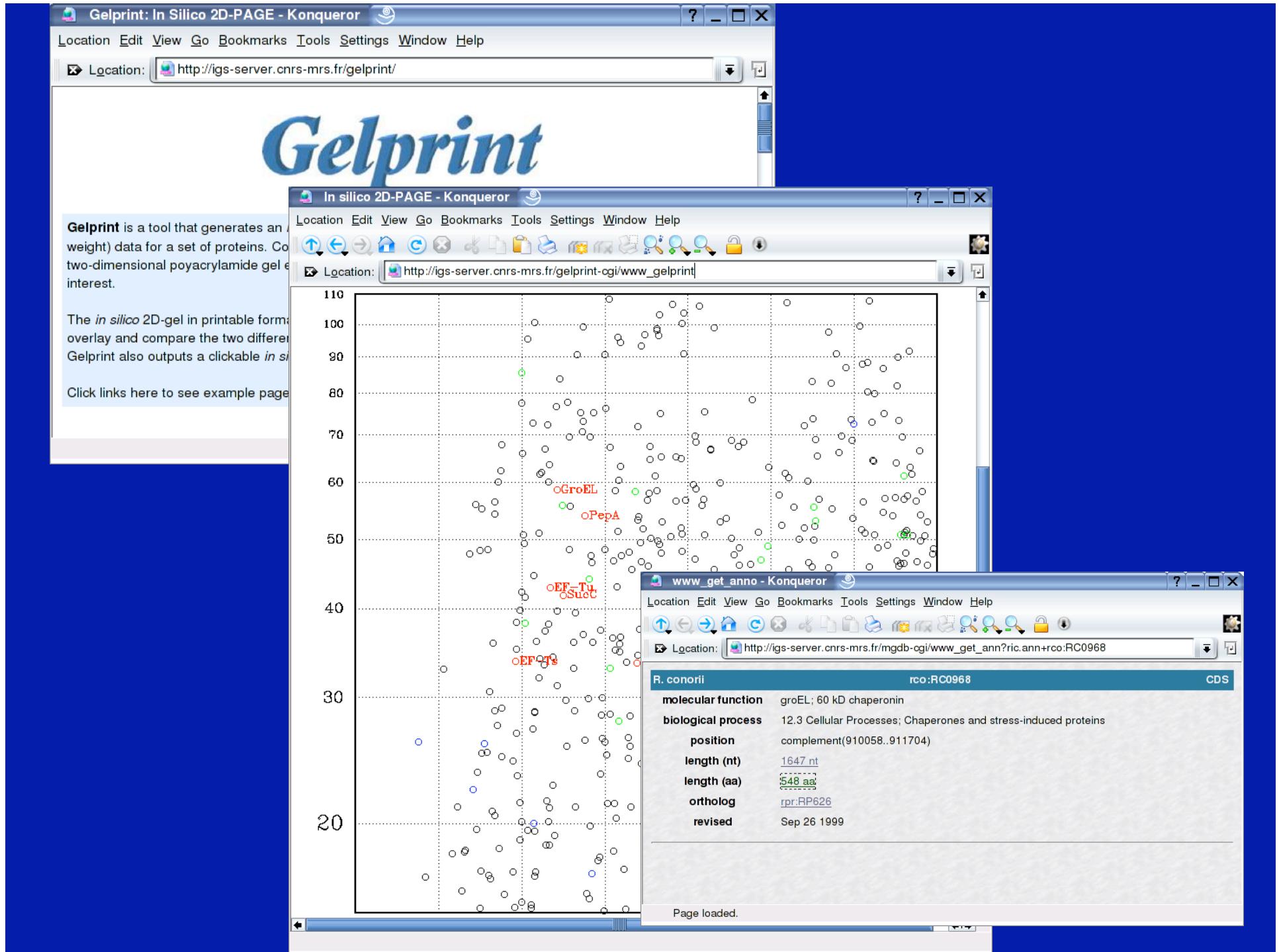
Sequest : 10 mn

Pour des résultats similaires

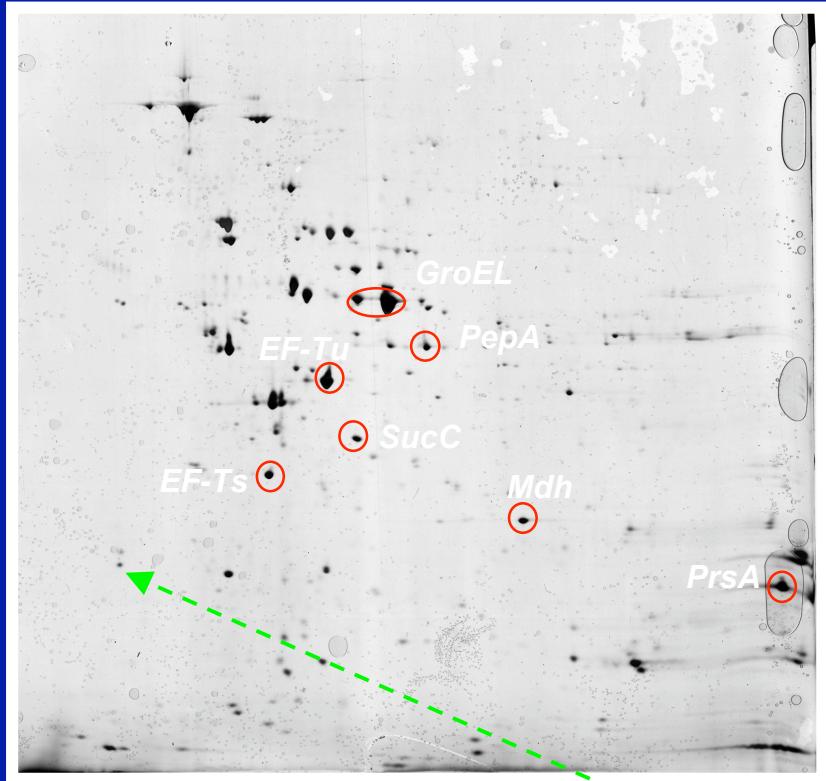
## 2D-gel of *Rickettsia conorii*



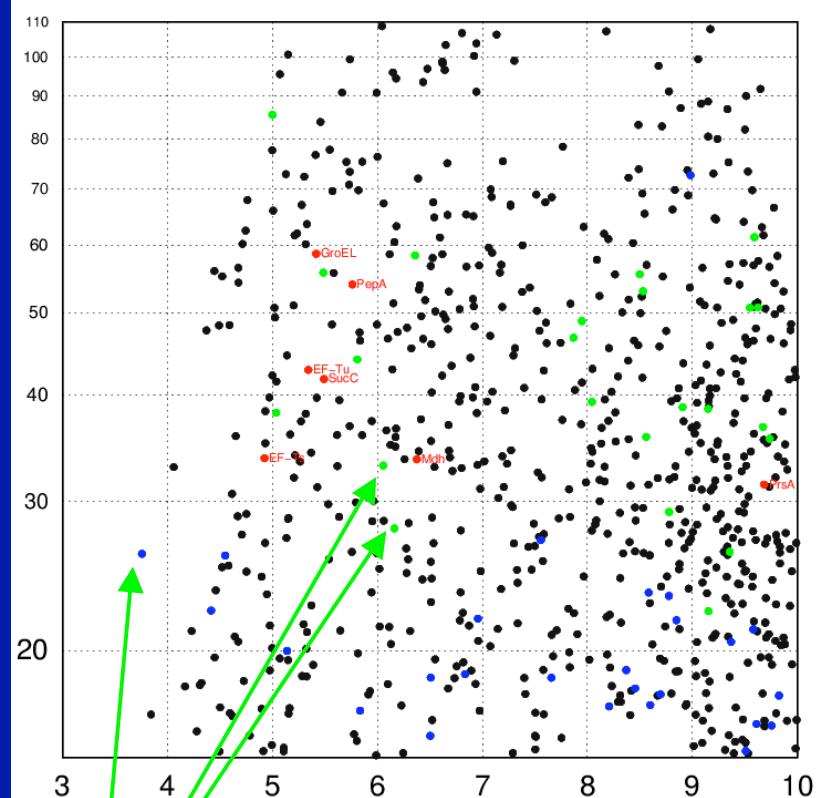
2D-gel provided by P. Renest et al.



## 2D gel (experimental)



## 2D gel (*in silico*)



?

**Proteins of interest !**  
“Split genes and Repeat-containing proteins”

## Input data for Gelprint

- List of theoretical (pre-computed) pI and Mw for a set of proteins
- Ranges of pI/Mw to show in the in silico 2D-gel
- Database address to make links from in silico 2D-gel to the database
- Gelprint generates in silico 2D-gels in Postscript/PDF that fit to your “real” gel
  - Size of your 2D-gel (in centimeter)
  - Some references points for fitting (i.e. positions of some identified spots on the real 2D-gel (in centimeter))

## Gelprint

<http://igs-server.cnrs-mrs.fr/gelprint/>

## Acknowledgements

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